

Optimization of Colour Reduction in the Pharmaceutical Effluent by Response Surface Methodology

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Abstract: This research deals with the reduction of colour in the pharmaceutical effluent by *Treculia Africans* seed coat (TA) as a coagulant using Box behnken design (BBD) from the response surface methodology (RSM). The pharmaceutical effluent was subjected to physicochemical analysis to determine the level of pollution. The coagulant was characterized by Fourier transformed infrared (FTIR) and Scanning electron micrograph (SEM). Result of the physicochemical analysis of the pharmaceutical effluent showed that the colour of the effluent is purple and its pH (8.11), Hardness (176 mg/L), phosphate 10.22 mg/L and turbidity 560 mg/L) were found to be above WHO permissible limit of effluent disposal. BBD generated 17 experimental run in which coagulation-flocculation process was carried out. These experimental data were analyzed by analysis of variance (ANOVA) and was found to fit 2nd order polynomial model (quadratic equation). The plot of predicted versus actual data confirmed that the model describe explicitly the colour reduction efficiency. The process parameters such as coagulant dosages (100-500 mg/L), settling time (10-50 minutes) and pH (2-10) were optimized to get the best treatment condition for colour reduction efficiency. The optimum colour reduction efficiency is 64.36% at coagulant dosage of 252.32 mg/L, settling time of 25.31 minutes and pH of 2.89. The SEM image after treatment suggested that pollutant has been removed from the pharmaceutical effluent since there is change in the surface morphology of the coagulant while FTIR analysis result after treatment proposed removal and addition of bonds due to interaction between the colloid particles of the pharmaceutical effluent and the coagulant. Hence, *Treculia Africans* seed coat (TA) can serve as alternative coagulant for reduction of colour from Pharmaceutical effluent.

Keywords: Pharmaceutical Effluent, Coagulant, Response Surface Methodology, *Treculia Africans*, Optimization

1. Introduction

As the horizon of industrialization is expanding likewise the pollution that emerges from these industries thus this poses a great risk to the ecosystem. Pharmaceutical company is one of the major industries that make use of chemicals in the manufacturing process. Sometimes the effluent that is disposed end up in our rivers, lakes, dams and even on our lands. This effluent may contains high chemical oxygen demand, biological oxygen demand, phosphate, nitrate, chloride, total dissolve solids, total suspended solid, turbidity, colour and heavy metals.

These pose a great danger to aquatic organism, man and the soil because of the pollutants liberated from this effluent. Coagulation-flocculation process is one of the treatment techniques used to treat water and waste water. Studies have shown that this process is cheaper and simple to use [1-4]. Coagulation is the process in which destabilization of particles occurs by the reduction of the repulsive potential of the electrical double layer [5] and flocculation is the phase whereby destabilized particles, or particles formed during destabilization are induced to collect into aggregates. These particles that are destabilized are referred to as colloids and finely divided matter [6]. Coagulation-flocculation takes place in successive steps,

permitting particle collision and growth of floc followed by sedimentation [7-9]. Coagulant is any substance that when added to water or waste water destabilizes the colloid particles of that water. Stable colloids in water normally present negative charges all around their surface [10]. Coagulants tend to cause the neutralization of these charges, thus colloidal particles become unstable and settle because of gravitational force.

The conventional coagulants that are used are made of up of inorganic and synthetic chemicals which are creating more pollution instead of solving them, hence, the need for natural coagulants. Researchers have opened windows of environmentally benign coagulants that is very much effective when used to purify effluents or water, mention but few are; chitosan [11], *Phoenix dactylifera* seed [12], *Moringa oleifera* seeds [13], *Cicer arietinum*, and *Dolichos lablab* [14], Drumstick seeds, Tamarind seeds, Neem seeds, Banana peel and sweet potato [15], Dragon fruit foliage [16] and Jackfruit Seed Powder [17].

Treculia Africana is a tree species in the genus *Treculia* and family *Moraceae*. The trees can grow below 1,500 metres (4,900 ft) above sea. The fruits are hard and fibrous, can be the size of a volleyball and weight up to 8.5 kg. It is known in English as African breadfruit in countries like Nigeria, Tanzania, Zambia, Uganda, USA. In Malawi, it is called African boxwood. The Ibos in southern Nigeria called it 'ukwa'. The seeds are used for soup thickening and making bread fruit cakes, snacks and cookies [18, 19].

The aim of this study is to reduce colour in the pharmaceutical effluent using the seed coat of *Treculia africana* (TA), optimizing the coagulation-flocculation process using Response surface methodology by Box behnken design (BBD) and characterizing the pharmaceutical effluent and the TA coagulant.

2. Materials and Methods

2.1. Collection of Samples

The pharmaceutical effluent was collected from the waste channel of a pharmaceutical industry located at Awka. The effluent collected is purple in colour. While *Treculia africana* seed coat (TA) was obtained from Nkpor main

market along Onitsha – Enugu expressway, Nkpor, Idemili North Local Government Area, both in Anambra state.

2.2. Preparation of Coagulant

The coagulant, *Treculia Africana seed coat* (TA) was washed and sun dried for 3 days to remove moisture. It was ground to powder, sieved and store for use in dry container.

2.3. Physicochemical Analysis

Determination of total suspended solid (TSS), total hardness, chloride, phosphate, nitrate, sulphate chemical oxygen demand (COD), biological oxygen demand (BOD) and total dissolved solid (TDS), were carried out according to the standard method for the examination of water and wastewater [20, 21]. Electrical conductivity, turbidity and pH were determined using electrical conductivity meter Model DDS-307, Turbid meter Lab. Tech. model 038, pH meter Hanna model HI991300.

2.4. Characterization of the TA Coagulant

FTIR and SEM of the TA coagulant were done using standard methods and instruments [22].

2.5. Coagulation-Flocculation Experiment

The Coagulation-Flocculation experiment was monitored by three factors; settling time from 10 mins, 20 mins, 30 mins, 40 mins and 50 mins, pH of 2, 4, 6, 8 and 10 and TA dosage from 100 mg/L, 200 mg/L, 300 mg/L, 400 mg/L and 500 mg/L. This experiment was performed at room temperature, 100 mg of the TA was weighed into 250 mL beaker and 100 mL of the effluent at pH of 2 was added into it. pH of the effluent was adjusted using 1 M HCl and 1 M NaOH. The content of the beaker was stirred for 2 mins of rapid mixing (80 rpm) and 2 mins of slow mixing (20 rpm) with a magnetic stirrer and then left undisturbed for 50 mins. It was filtered and the filtrate was subjected to color reduction analysis using UV/VIS spectrophotometer at lambda (λ) maximum of 510 nm (Figure 1) to determine the absorbance and thus colour reduction efficiency is calculated as;

$$\% \text{ colour reduction efficiency} = \frac{\text{Initial absorbance} - \text{final absorbance}}{\text{Initial absorbance}} \times 100\%$$

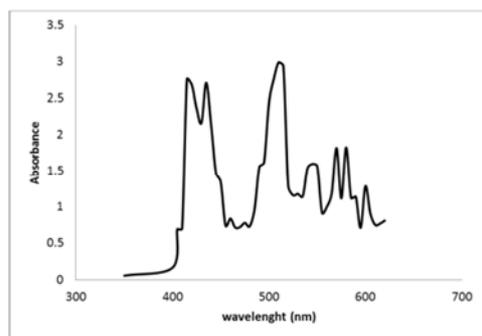


Figure 1. Plot of Absorbance versus wavelength showing the wavelength maximum (λ_{max}).

The experiment was repeated accordingly as written in Table 2 which is the design matrix of Box-behnken Design (BBD).

2.6. Response Surface Methodology

Design expert 11 was used to design the experiment. Here three-level Box-behnken experimental design with categorical factor was employed to study the synergistic effect of the various parameters namely, coagulant dosage (X_1), settling time (X_2) and pH (X_3) and also to optimize the coagulation-flocculation process of TA coagulant as shown in Table 1 and a total of 17 experimental runs were obtained. The experimental design matrix by the Box-Behnken design showing the interaction of the independent variables/factors

with response as the colour reduction efficiency is tabulated in Table 2. The response variable fitted to 2nd order polynomial (quadratic equation) model as shown in equation (1):

$$\gamma = \beta_0 + \sum_{i=1}^k \beta_i X_i + \sum_{i=1}^k \beta_{ii} X_i^2 + \sum_{i < j} \beta_{ij} X_i X_j$$

Where γ is the response (colour reduction efficiency) variable to be modeled; X_i and X_j the independent variables

which influence γ ; β_0 , β_i , β_{ii} and β_{ij} are the offset terms, the i th linear coefficient, the quadratic coefficient and the ij th interaction coefficient, respectively [23].

Table 1. Factors and levels used in BBD.

Factors	Lower level	Medium level	High level
Coagulant dosage (X_1)	100 mg/L	300 mg/L	500 mg/L
Settling time (X_2)	10 minutes	30 minutes	50 minutes
pH (X_3)	2	6	10

Table 2. Design Matrix of Box behnken design.

Run	Factor 1 X_1 : Dosage (mg/L)	Factor 2 X_2 : Settling time (minutes)	Factor 3 X_3 : pH	Colour reduction Efficiency (%)
1	300	30	6	51.43
2	500	30	10	35.79
3	300	50	10	17.21
4	100	30	2	52
5	500	30	2	58.82
6	100	50	6	20.87
7	300	50	2	59.14
8	300	10	2	68.45
9	500	50	6	38.68
10	300	10	10	24.22
11	100	10	6	33
12	500	10	6	47.86
13	300	30	6	51.9
14	300	30	6	51
15	300	30	6	51.43
16	300	30	6	51.43
17	100	30	10	10.5

3. Results and Discussions

3.1. Physicochemical Result of the Pharmaceutical Effluent

Physicochemical result obtained in Table 3 shows that the colour of the effluent is purple thus need for the removal of the colour since colour from pharmaceutical effluent came from colourants added and components mixtures during production of drugs and it has high thermal and photo

stability to resist biodegradation when discharged to the environment. Since the final route of effluent is lake, sea, river, dam etc, too much accumulation of this colours on the water bodies can stop the re-oxygenation capacity of the receiving water thus cut-off sunlight, thereby upsetting biological activity in aquatic life and also the photosynthesis process of aquatic plants or algae [24, 25]. From the same Table 3 it can be observed that Hardness, Phosphate and turbidity are all above the standard limit proposed by WHO.

Table 3. Physicochemical result of the pharmaceutical effluent.

Parameters	Values	WHO Standard
Colour	Purpleabsorbance=5.073	
pH	8.11	6.5 – 8.5
TSS (mg/L)	0.16	500
TDS (mg/L)	0.36	500
COD (mg/L)	66.6	250
BOD (mg/L)	172	500
Alkalinity (mg/L)	60	200
Acidity (mg/L)	0.0256	200
Hardness (mg/CaCO ₃)	176	70
Nitrate (mg/L)	9.19	40
Sulphate (mg/L)	169.95	500
Phosphate (mg/L)	10.022	3.5 – 5.0
Chloride (mg/L)	37	250
Turbidity (NTU)	560	5 – 10
Temperature (°C)	31.3	30-35
Conductivity (µs/cm)	179.0	500

3.2. Statistical Analysis from BBD

The statistical significance of the quadratic model was

evaluated by the analysis of variance (ANOVA), as presented in Tables 4. The quadratic equation obtained is given in equation 1:

$$Y = 51.44 + 8.1X_1 - 4.7X_2 - 18.84X_3 + 0.7373X_1X_2 + 4.62X_1X_3 + 0.525X_2X_3 - 9.66X_1^2 - 6.68X_2^3 - 2.5X_3^2 \quad (1)$$

Model terms were evaluated by the p value, lack of fit, F test, coefficient of determination (R^2) and adequate precision (AP). The term with p values more than 0.05 are insignificant while less than 0.05 is significant (Table 4). In order to develop the regression model that is statistically significant, the insignificant terms in equation 1 as shown in Table 4 were eliminated. The insignificant

terms in equation 1 are X_1X_2 , X_2X_3 and X_3^2 . These terms have p values more than 0.05 hence are removed to obtain the regression model equation 2. Positive sign in front of these equations indicate synergistic effect of the factors, whereas negative sign indicates antagonistic factor effect [26].

$$Y = 51.44 + 8.1X_1 - 4.7X_2 - 18.84X_3 + 4.62X_1X_3 - 9.66X_1^2 - 6.68X_2^3 \quad (2)$$

P values < 0.05 for colour reduction demonstrate that the model for those response variables were significant at the 95% confidence level [26]. F test-value shows that the model is significant. In addition the model exhibited significant lack of fit. Therefore, what measures the overall performance of a model is R^2 . When predicted R^2 and adjusted R^2 differ dramatically, there is a good chance that non-significant terms have been included in the model [27]. The predicted R^2 is 0.7731 and is in reasonable agreement with the adjusted R^2 of 0.9857 because their difference is less than 0.2.

A reliable model should also be able to predict response with a reasonable accuracy when compared with the experimental data. Figures 2 is the plot of Predicted versus Actual data for TA coagulants. It compares experimental colour reduction efficiencies (%) with the predicted values obtained from the models. The observed points on these plots revealed that the actual values are distributed comparatively near to the straight line, indicating that the regression model is able to predict these colour reduction efficiencies [28-30]. Thus Figure 2 indicated good agreement between the experimental and predicted values.

DESIGN-EXPERT Plot
Response

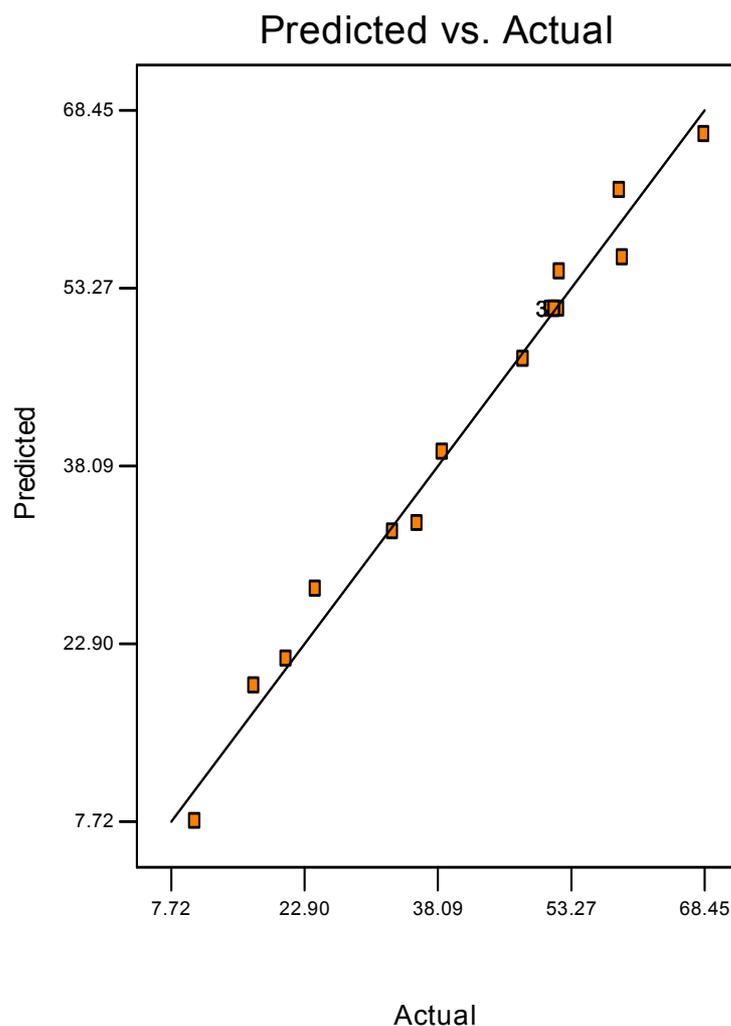


Figure 2. A plot of Predicted against actual data.

DESIGN-EXPERT Plot

Response
X = A: Dosage
Y = B: Settling time

Actual Factor
C: pH = 6.00

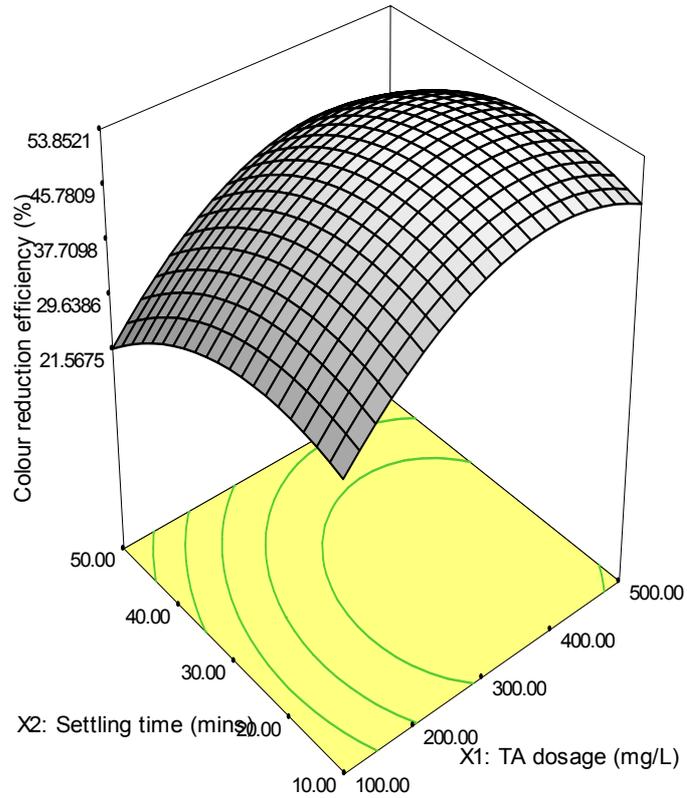


Figure 3. 3D surface interactive effect between Coagulant dosage (TA) and settling time (X_1 & X_2) at middle level pH 6.

DESIGN-EXPERT Plot

Response
X = A: Dosage
Y = C: pH

Actual Factor
B: Settling time = 30.00

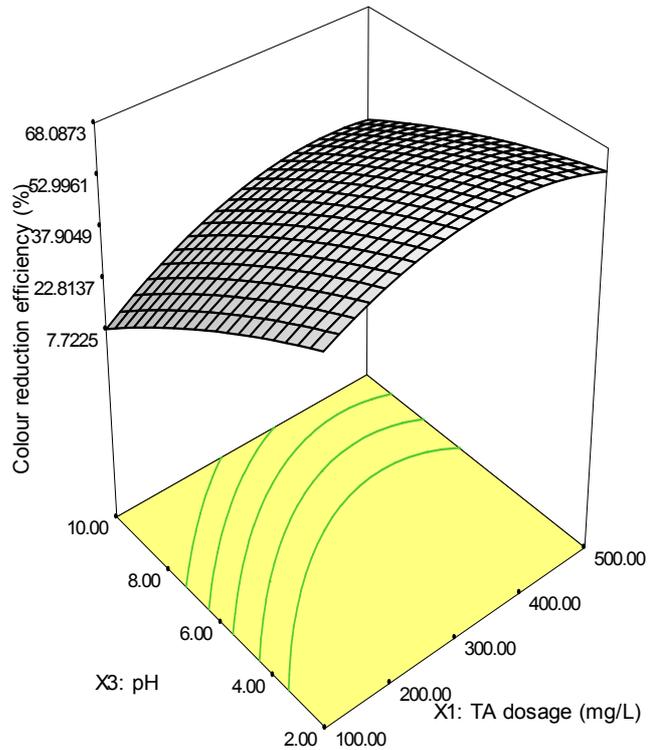


Figure 4. 3D surface interactive effect between coagulant dosage (TA) and pH (X_1 & X_3) at middle settling time of 30 mins.

DESIGN-EXPERT Plot

Response
 X = B: Settling time
 Y = C: pH
 Actual Factor
 A: Dosage = 300.00

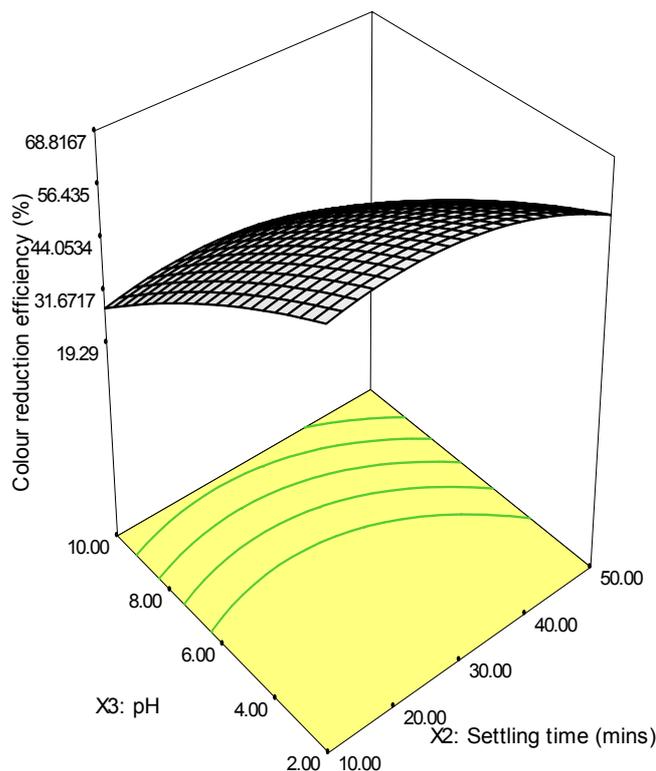


Figure 5. 3D surface interactive effect between settling time and pH (X_2 & X_3) at middle coagulant dosage of 300 g/mL.

Table 4. ANOVA for Response Surface Quadratic model for MMM coagulant.

Source	Sum of Squares	df	Mean Square	F-value	p-value	
Model	4288.8	9	476.53	53.77	< 0.0001	significant
X_1 : Dosage	524.56	1	524.56	59.19	0.0001	
X_2 : Settling time	177	1	177	19.97	0.0029	
X_3 : pH	2838.43	1	2838.43	320.27	< 0.0001	
X_1X_2	2.18	1	2.18	0.2455	0.6354	
X_1X_3	85.29	1	85.29	9.62	0.0173	
X_2X_3	1.32	1	1.32	0.1492	0.7108	
X_1^2	392.62	1	392.62	44.3	0.0003	
X_2^2	187.83	1	187.83	21.19	0.0025	
X_3^2	26.4	1	26.4	2.98	0.128	
Residual	62.04	7	8.86			
Lack of Fit	61.63	3	20.54	202.67	< 0.0001	significant
Pure Error	0.4055	4	0.1014			
Cor Total	4350.84	16				
R^2	0.9857					
Adjusted R^2	0.9674					
Predicted R^2	0.7732					
Adeq Precision	25.6810					

3.3. Interactive Effect of the Three Independent Parameters

The interactive effect of X_1 , X_2 and X_3 were shown on response surface plots, where some variable were kept constant at middle level in Figures 3-5. The response surface plots indicate that the maximum colour removal efficiencies are located inside the design boundary. In Figures 3, 4, and 5, it can be observed that, the colour reduces at the condition where TA dosage is within the range of 200-400 mg/L, and pH 8-2 and settling time of 20-40 mins recorded a little

increase in the colour reduction efficiency. TA dosage and pH in Figures 3, 4 and 5 plays a great part in the colour reduction efficiency. Therefore, TA dosage, pH and settling time are the significant factors for colour reduction efficiency. Settling time showed very little effect on the colour reduction efficiency and this can be attributed to the coagulation-flocculation hydrolysis, which can inhibit the bridging and enmeshment [30]. The mechanism that can be proposed in this coagulation-flocculation process is charge neutralization. The existence of different charges induces mutual attraction

between particles to spontaneously agglomerate since repulsion force were overcome resulting in an increase in the stability of the suspension. Hence in acidic condition and adequate dosage an increase in the colour reduction efficiency is observed. Where there is an excess TA dosage charge reversal is favoured thus, decrease in the colour reduction efficiency. This result aligned with the result obtained elsewhere [31, 32].

3.4. Optimization of the Coagulation-flocculation for TA Coagulant

Numerical optimization was applied from BBD design to obtain a desirable value, optimum process parameters and to achieve highest treatment performance for colour reduction efficiency of TA coagulant used for each input factors and

response can be selected. Equation 2 was solved for the best solutions such that the responses γ are maximized within the design space. The desirability of 1.00 and optimum condition for colour reduction efficiency is 64.36% at coagulant dosage of 252.32 mg/L, settling time of 25.31 minutes and pH of 2.89 were obtained.

To verify the reliability of the regression model, the pharmaceutical effluent was retreated at the optimum conditions in triplicate. The colour reduction efficiency obtained through the experiment is $63.66 \pm 0.42\%$. This result is close to the predicted value estimated by the BBD, thus this confirms that the RSM approach is suitable for optimizing the operational conditions of the coagulation-flocculation process.

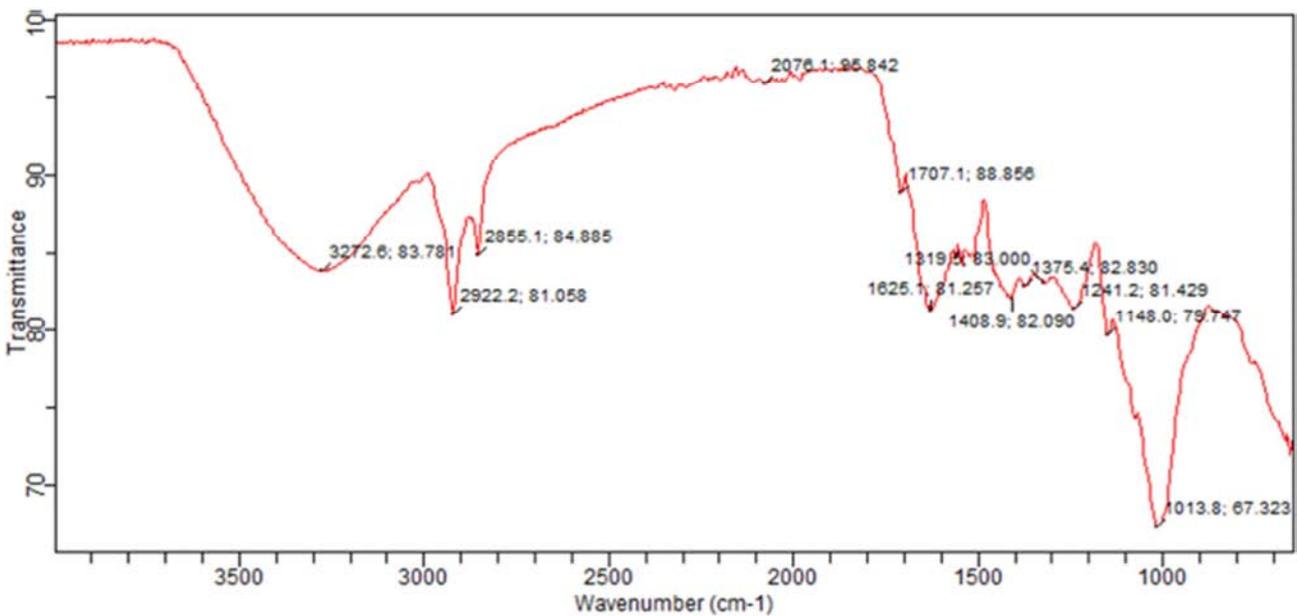


Figure 6. FTIR Spectrum of TA before coagulation- flocculation process.

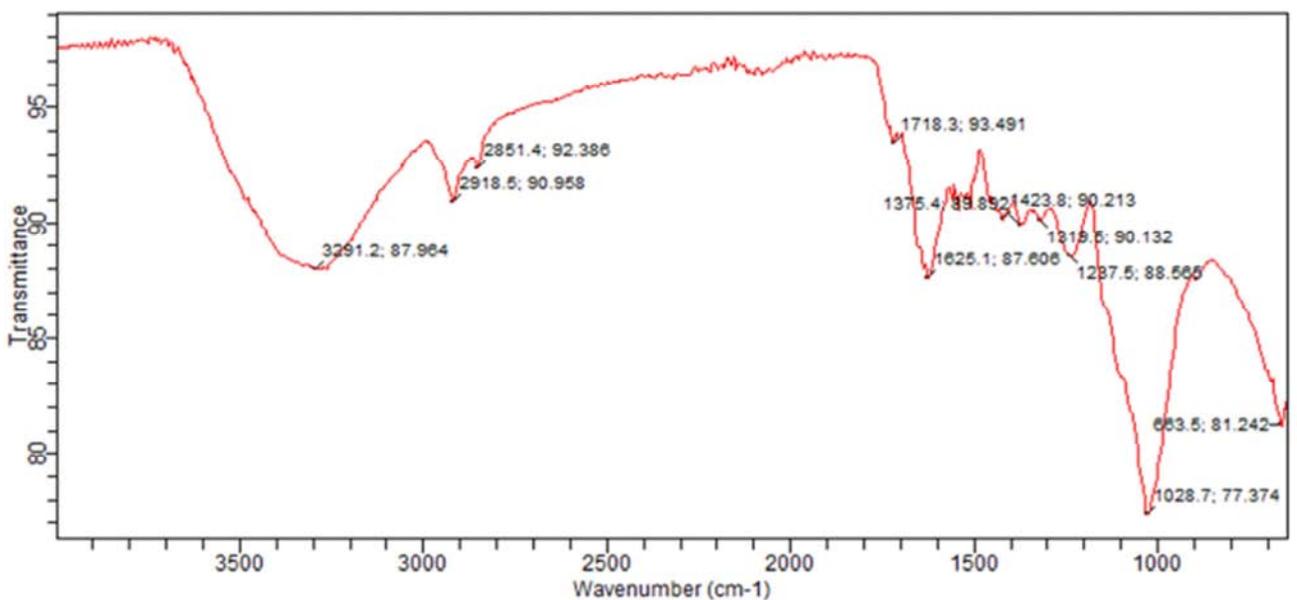


Figure 7. FTIR TA after coagulation- flocculation process.

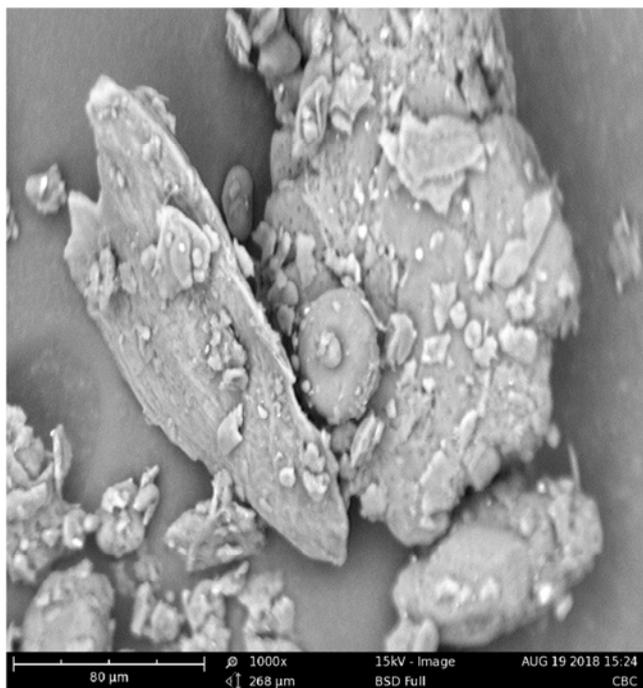


Figure 8. SEM image of TA before Coagulation-flocculation process.

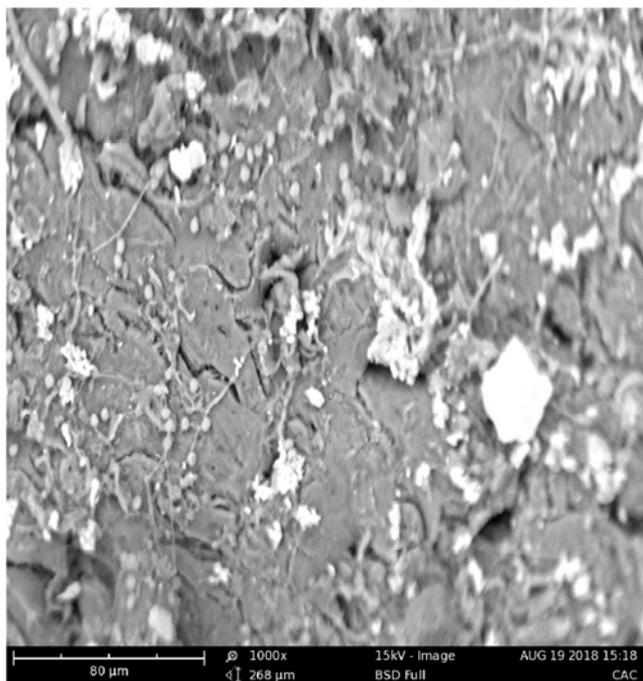


Figure 9. SEM image of TA after Coagulation-flocculation process.

3.5. Characterization of the TA Coagulant

FTIR spectra of the TA before and after coagulation-flocculation process are shown in Figures 6 and 7. In Figure 6, the peak at 3292.6 cm^{-1} is an indication of O-H stretch from alcohol, Absorption bands at 2922.2 and 2855.1 cm^{-1} are C-H stretch from alkane. Bands at 2076.1 cm^{-1} and 1707.1 cm^{-1} are N=C=S stretch, 1625.1 and 1408.9 cm^{-1} are O-H bending. The peaks at 1375.4 , 1319.5 , 1241.2 and 1148.0 cm^{-1} are C-O stretch and band at 1013.8 cm^{-1} is referred to as C=C bend.

While Figure 7, displays broad and intense peaks as seen in Figure 6 but with some modifications as a result of the interaction of the pollutant including the colour on the surface of the TA coagulant. Absorption band at 3291.2 cm^{-1} demonstrated O-H stretch, 2918.5 and 2851.4 cm^{-1} are C-H stretch, 1718.3 , 1625.1 , 1423.8 , 1319.5 and 1237.5 cm^{-1} are O-H bend. The peak at 1028.7 cm^{-1} is C-O stretch. Comparing Figures 6 and 7, it can be inferred that after the coagulation-flocculation process, there were shift and removal of the absorption bands in the spectral bands indicating an interaction between the colloid particles of the pharmaceutical effluent and that of the TA coagulant hence the shifting and removal of some functional groups.

SEM image of TA coagulant before and after coagulation-flocculation process are shown in Figures 8 and 9. In Figure 8, the SEM image shows that the coagulant has rough edges and vacant spaces which is called pores while in Figure 9, the pores and edges has been occupied by the colloid particles of the pharmaceutical effluent. The change in the surface morphology of the coagulant after the treatment indicates the adsorption of pollutants on the surface of the coagulant hence TA can be used as a bio-coagulant in the colour reduction of the pharmaceutical effluent.

4. Conclusion

The study has shown that response surface methodology by BBD optimized the colour reduction efficiency. ANOVA report stated that the 2nd order polynomial (quadratic) equation is the best model for this process. All the independent variables tested such as coagulation dosage, settling time and pH showed significant interaction and contributed to the increase in the colour reduction efficiency. The best pH condition was found to be at acidic medium. From the result of FTIR and SEM analysis it can be suggested that what brought about the reduction of the purple colour of the pharmaceutical effluent treated with TA coagulant is the surface chemistry of the TA coagulant.

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