

Research Article

Genetic Characterization of Populations of *Zeugodacus Cucurbitae* (Coquillet, 1899), a Watermelon Pest

Madeleine Ivonne Mendy* , Toff ène Diome , Mamecor Faye ,
Mback éSemb ène 

Department of Animal Biology, Cheikh Anta Diop University, Dakar, Senegal

Abstract

Zeugodacus cucurbitae or melon fly formerly called *Bactrocera cucurbitae* is an agricultural pest of Asian origin. Well known as a pest of fleshy fruits and vegetables damaging 81 host plants, the melon fly has been the subject of several studies due to its introduction and dissemination worldwide. Up to now, no study on the global structuring of *Zeugodacus cucurbitae* has been done. Therefore, knowledge of its genetic structuring would allow better management of the insect. It is in this context that the present study on the genetic characterization of populations of *Z. cucurbitae* watermelon pest insect fits. Our data was collected from the Genbank database. Phylogeographic analyses were made using mitochondrial cytochrome oxidase I (COI) DNA as a genetic marker. After analysis, the study demonstrated two distinct groups: a group composed of the population of Reunion and another group composed of populations from Africa, Asia, Oceania, and Hawaii. This is the result of a genetic isolation demonstrated by the Mantel Test for which the significant p-value confirms the correlation between genetic distances and geographical distances. However, there is a genetic differentiation between individuals in the Reunion population. For any fight against this insect, it would be interesting to take into account the existence of these two genetic groups.

Keywords

Zeugodacus Cucurbitae, Characterisation, COI, Phylogeography, Mitochondrial DNA

1. Introduction

Agriculture is a pillar of sustainable development, economic stability and food security for a country. Nowadays, watermelon is considered a "functional food" and a popular fruit with important nutritional and bioactive compounds offering several health benefits [1]. The watermelon belongs to the Cucurbitaceae family. According to Badji [2], cucurbits such as melon, watermelon, and cucumber are also crops that are of increasing interest for export needs. Despite these assets, these crops are threatened by pest insects. *Zeugodacus cucurbitae*, also called the watermelon fly, is a serious pest

belonging to the family Tephritidae. It is a major pest of fleshy fruits and vegetables [3]. It has economic importance due to the damage it causes to vegetable and fruit crops [4, 5]. Probably originating from India [6], this species is very widespread throughout the world [7-10]. Understanding the genetic structure and the genetic diversity of the insect is crucial for effective management. Previous studies have been conducted using cyto-chromium oxidase I in China, Southeast Asia [11], India [12], East Asia, Central Asia, Africa, Hawaii and Reunion [8] on the genetic relationships between popu-

*Corresponding author: madeleineivonne.mendy@ucad.edu.sn (Madeleine Ivonne Mendy)

Received: 19 August 2025; **Accepted:** 3 September 2025; **Published:** 14 October 2025



Copyright: © The Author(s), 2025. Published by Science Publishing Group. This is an **Open Access** article, distributed under the terms of the Creative Commons Attribution 4.0 License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited.

lations and their demographic evolution. They revealed low genetic diversity between populations and a genetic structuring according to environmental zones. Therefore, an assessment of the degree of homogeneity at the global level would be advantageous for the control of this pest.

It is in this wake that this study is undertaken, thus aiming to understand the genetics of *Zeugodacus cucurbitae* populations.

2. Materials and Methods

2.1. Study Sites

Our study took place in a total of 20 countries spread over the 5 continents:

- 1) Africa: Senegal, Tanzania
- 2) Oceania: Guam, Northern Mariana Islands, Solomon Islands
- 3) America: Hawaii
- 4) Europe: Reunion
- 5) Asia: Thailand, Japan, Laos, Cambodia, Nepal, Bangladesh, Indonesia, Malaysia, China, Sri Lanka, Taiwan, Philippines, Vietnam

2.2. Study Populations

The study populations were defined according to countries. Individuals from each country constitute a population. We have a total of 20 populations.

2.3. Data Collection

2.3.1. Interest in Cytochrome Oxidase I

The study was carried out on a molecular marker of mitochondrial DNA: cytochrome oxidase I (COI) which codes for a subunit of a transmembrane protein complex of the respiratory chain. Mitochondrial genes have low diversity among individuals of the same species because mitochondria are transmitted vertically from mother to daughter, thus creating a bottleneck [13]. The COI is widely used in studies of phylogeny, phylogeography and population genetics but also for species identification [14-20]. It is a determining marker in the process of Barcoding [15-18]. Its rate of molecular evolution is rapid in addition to the absence of meiotic recombination (generally), variations are the result of accumulations of mutations allowing to consider polymorphic sequences as unique haplotypes [19].

In other words, the mitochondrial genome of animals is a better target for analysis than the nuclear genome due to its lack of introns, limited exposure to recombination, and haploid inheritance pattern [21].

2.3.2. Acquisition of Sequences

Sequences were downloaded from GENBANK genomic databases. It is a database that collects DNA sequences

available publicly on NCBI (National Centre for Biotechnology Information: www.ncbi.nlm.nih.gov/genbank). Sequences are removed from Genbank thanks to their accession number and downloaded under the FASTA extension (fasta). They were then grouped by continent with 120 sequences from Asian countries (Malaysia, Thailand, Bangladesh, Sri Lanka, Vietnam, Indonesia, Philippines, China, Japan, Laos, Cambodia, Nepal, Taiwan), 25 sequences from Oceania countries (Solomon Islands, Northern Mariana Islands, Guam), 20 sequences from the countries of Africa (Senegal, Tanzania), 10 sequences from the USA (Hawaii) and 10 sequences from Europe (Reunion).

2.4. Genetic Analyses

2.4.1. Sequences Analysis

(i). Sequences Alignment

Sequences alignment is the first step in phylogenetic analysis. It is important because it allows us to evaluate the homology of the sites. Our sequences have been aligned and corrected by the BioEdit software version 7.2.5 [22] using the ClustalW algorithm. As for the cleaning of the sequences, it is done in conjunction with the alignment.

(ii). Basic Parameters of Genetic Diversity

As part of the sequence polymorphism analysis, the DnaSP software version 5.10.01 [23] was used to determine the identity card of our data, that is, the basic parameters of variability which are the number of sites (N), the sample size (n), variable sites such as singleton or non-informative sites and informative sites in parsimony. The MEGA 7.0.14 [24] software was used to extract the nucleotidic frequencies which are the nature of the mutations, the rate of mutations (R) and the type of substitutions i.e. synonymous (Ks) and non-synonymous (Kns) mutations. The same software made it possible to highlight the percentages of transitions and transversions calculated using the nucleotide substitutions model.

2.4.2. Genetic Diversity Indices

The DnaSP software version 5.10.01 [23] made it possible to determine genetic diversity indices, which are nucleotide diversity π and haplotype diversity (Hd), as well as their variances. Moreover, π is defined as the probability according to which two sequences randomly drawn in a sample are different at a given site and Hd the probability that, two haplotypes randomly drawn in a population are similar [25]. We have the signal of a stable population with a large effective size (or admixture) when Hd and π are strong; When the latter are weak, we have the signal of a severe and prolonged demographic bottleneck. However, a strong Hd and a weak π would mean rapid population growth from an ancestral population with low numbers and where time is not sufficient to find a high diversity between haplotypes. A

short-lived bottleneck in a large ancestral population would mean low Hd and high Pi [26].

2.4.3. Haplotype Networks

Haplotype networks are an application of the median link model to show the relationships between different haplotypes. A minimum haplotype network is characterized by nodes (circles) and branches (links) that connect the nodes. Each node corresponds to a haplotype whose size is proportional to the frequency of the haplotype in the dataset [16]. They are built with NETWORK version 10.2 using the Median-Joining method [27] in order to identify their phylogenetic relationships.

2.4.4. Differentiation and Genetic Distance

Structuring requires the parameters of genetic differentiation, which are the F_{st} (the degree of genetic differentiation) determined with the software Arlequin version 3.5.1.3 [28] and the genetic distance D executable with the program MEGA 7.0.14 [24]. The genetic distance is calculated within each population (intra-populations distances) and between populations taken two by two (inter-populations distances).

2.4.5. Structuration

(i). Correlation Test Between Genetic Distance and Geographical Distance

Mantel test was used to measure the influence of geographical distance on genetic differentiation thanks to XLSTAT software version 2021.5. Geographical distances were obtained from Google earth. The hypothesis that genetic and geographical distances are not correlated is null. The hypothesis that genetic and geographical distances are correlated is alternative. If the p-value is below the threshold significance level of 0.05, the null hypothesis is rejected. Otherwise, the null hypothesis is accepted.

(ii). AMOVA Test

The Analysis of Molecular Variance (AMOVA) implemented in the software Arlequin version 3.5.1.3 [28] allows to determinate the genetic structuring of populations based on a given factor.

2.4.6. Demographic Evolution

The genetic demo tests were carried out in order to distinguish the sequences (loci) whose evolution follows a neutral model to those evolving according to a non-random process. It is a question of comparing the level of adjustment between the diversity observed at the loci and that expected under the hypothesis of a neutralist model (at the mutation-drift equilibrium).

The estimation of Tajima's D , Fu's and Li's F_s was done with the software Arlequin [28]. These values test the hypothesis that mutations are selectively neutral. The qualitative graphical representation of the distribution of genetic distances between individuals in a population taken two by two was done using the Dnasp software version 5.10.01 [23].

2.4.7. Phylogenetic Approach

Phylogenetic analysis is one of the main tools of evolutionary biology. Indeed, it allows the evaluation of the kinship relationships between the different strains of *Z. cucurbitae*. The reconstruction of phylogenetic trees was carried out using different approaches:

- 1) Distance methods: Neighbor joining (Kimura 2-parameter) which constructs the tree based on the similarities observed between each pair of sequences. It is based on the matrix of genetic commands.
- 2) Character methods: The maximum likelihood that evaluates, in terms of probabilities, the order of branches and the length of the branches of a tree within the framework of a given probability evolution model. It allows you to see all the events that may have generated the current data set analysed. This method leads to the result closest to the real evolutionary tree. The Akaike information criterion (AIC) was used to choose the model for the evolution of the sequence. The best model is the one with the lowest AIC value. In our case, the latter corresponds to model TN93 + G + I. The latter will be retained for the reconstruction of the trees. These trees are verified with the MEGA software version 7.0.14 [24] and the robustness of the branches has been evaluated at 1000 Bootstrap.
- 3) The Bayesian approach is evaluated thanks to the skylog Mr BAYES version 3.2.6 [29]. The distribution of posterior probabilities of trees was estimated by 4 MCMC chains (3 of which were burned gradually and one cold). One million (1,000,000) generations were carried out for each of the chains by sampling the different parameters of all the 1,000 generations. The degree of convergence of the chains can be verified by examining the evolution of the likelihood function during the journey of the "cold" chain in order to determine the ignition period. The generations carried out during this period are eliminated from the analyses. In a conservative manner, the first 250,000 generations were eliminated (25%) and the inferences were then made on the next 750,000 generations. The model used is the same as that of the likelihood (TN93 + G). The visualization of the trees was done with Fig Tree version 1.4.2 [30]. A sequence of *Bactrocera dorsalis* was used as an out-group for the construction of phylogeographic trees.

3. Results

3.1. Genetic Variability

3.1.1. Basic Parameters of Genetic Variability

After alignment and cleaning, our dataset presents a total of 182 nucleotide sequences with each 592 sites including 33

sites with gaps. Among these sites, 247 are monomorphic and 312 are polymorphic including 12 singleton sites and 300 informative variable sites. The singleton sites have 2 variants and are at the following positions: 33, 117, 132, 254, 324, 338, 359, 371, 389, 429, 542, and 549. Within the informative variable sites, 284 sites constitute 2-variant sites, 13 sites are 3-variants and 3 sites are 4-variants. Moreover, the total number of mutations (Eta) is 331. All these results are summarized in [Table 1](#).

Table 1. Genetic diversity parameters.

E	N	M	I	S=S2V	P	P2V	P3V	P4V	Eta
182	592	247	312	12	300	284	13	3	331

E: sample size; N: total number of sites; M: monomorphic site names; I: polymorphic site names; S: singleton site names; S2V: singleton site names with 2 variants; P: information site names; P2V: information site names with 2 variants; P3V: name of information sites with 3 variants; P4V: name of information sites with 4 variants; Eta: total number of mutations.

On all mutations, transitions (69.94%) are more numerous than transversions (30.06%). Transitions between pyrimidine bases (35.58%) are slightly more numerous than those between purine bases (34.36%). For transition-type mutations, in 24.20% of mutations, cytosine replaces thymine and vice versa for 11.38% of mutations. In 22.30% of mutations, guanine replaces adenine and the latter replaces the former in 12.06% of mutations. For those of the transversion type, adenine and guanine replace thymine (5.79%) and cytosine (2.72%). Similarly, the latter replace adenine in 4.23% and guanine in 2.29%. All these results are recorded in [Table 2](#).

Table 2. Percentage of Transition and transversion.

Bases	A	T	C	G
A	-	5.79	2.72	12.06
T	4.23	-	11.38	2.29

Bases	A	T	C	G
C	4.23	24.20	-	2.29
G	22.30	5.79	2.72	-

The average number of nucleotide differences between species (K) was deduced according to the following two groups: Country (all populations except Reunion) and island (Reunion). The group of countries (K=0.784) is more homogeneous than Reunion Island (K=1.756).

3.1.2. Genetic Diversity Indices

Nucleotidic (Pi) and haplotypic (Hd) diversities are determined ([Table 3](#)) within each population and for all populations. For all populations, the Hd and Pi values are high. Populations like Bangladesh, Sri Lanka, Vietnam, and Nepal all have high Hd and low Pi. All other populations have low Hd and low Pi.

Table 3. Haplotypic (Hd) and Nucleotidic (Pi) diversities of populations.

	Total population	Senegal	Tanzania	Thailand	Bangladesh	Sri Lanka	Indonesia
hd	0.390+/-0.002	0.2+/- 0.238	0.378+/-0.032	0.378+/-0.032	0.778+/-0.0188	0.778+/-0.02	0.286+/-0.038
Pi	0.056+/- 0.102	0.000+/-0.001	0.001+/-0.001	0.001+/-0.001	0.0017+/-0.003	0.003+/-0.004	0.0005+/-0.001
	China	Vietnam	Philippines	Taiwan	Laos	Malaysia	Nepal
hd	0.378+/- 0.033	0.778+/-0.019	0.200+/-0.024	0.378+/-0.033	0.200+/-0.024	0.222+/-0.028	0.833+/-0.016
Pi	0.004+/-0.006	0.002+/-0.0036	0.000+/-0.0006	0.000+/-0.0012	0.000+/-0.0006	0.000+/-0.0006	0.003+/-0.0044
	Japan	Mariana Islands	Salomon Island	Guam	Hawaii	Reunion	Cambodia

	Total population	Senegal	Tanzania	Thailand	Bangladesh	Sri Lanka	Indonesia
hd	0.400+/-0.0563	0.417+/-0.036	0.0	0.00	0.200+/-0.023	0.378+/-0.0328	0.286+/-0.0385
Pi	0.0006+/-0.0008	0.000+/-0.001	0.0	0.00	0.00034+/-0.00060	0.00312+/-0.00503	0.00049+/-0.00069

3.1.3. Haplotype Networks

Twenty-seven haplotypes were enumerated for our dataset. The haplotypes H1, H4, H5, H8, H14, and H26 are common. H1 is the majority haplotype with 142 individuals representing 78.02% of the 182 sequences and is shared in all sampled localities except Reunion. Haplotype 26 is a private haplotype consisting of 8 animals all from Reunion. Haplotype 8 is a common haplotype containing 4 individuals including 1 from Sri Lanka, 1 from Indonesia, 1 from China and 1 from the Philippines. Common haplotype 14 consists of 3 individuals from Vietnam, Taiwan, and Malaysia. The haplotypes 4 and 5 each have 2 individuals: haplotype 4 with Bangladesh and Laos and haplotype 5 with Bangladesh and Nepal. The individual haplotypes are 21 in number, including 4 from Sri Lanka (Hap_9, 10, 11 and 12), 4 from Nepal (Hap_18, 19, 20 and 21), 2 from Bangladesh (Hap_6 and Hap_7), 2 from

Vietnam (Hap_15 and Hap_16), 2 from Reunion (Hap_25 and Hap_27), 1 from Hawaii (Hap_24), 1 from Taiwan (Hap_17), 1 from China (Hap_13), 1 from Thailand (Hap_3), 1 from Tanzania (Hap_2), 1 from Cambodia (Hap_22) and 1 from the Northern Mariana Islands (Hap_23). In the network (Figure 1), some haplotypes are regressed into haplogroups (SN1 and SL2). Haplogroup SN1 includes the haplotypes H1, H2, H3, H4, H5, H6, H7, H11, H12, H14, H15, H17, H19, H20, H21, H22, H23, and H24. In the SL2 haplogroup (8 individuals) are concentrated the haplotypes H8, H9, H10, H16, and H18. The SN1 haplogroup is distributed in all sampled countries except the Reunion; it is composed of 163 individuals, or 89.56% of the total population. The SL2 haplogroup with 2.19% of the total population is composed mainly of individuals coming from Sri Lanka. The haplotypes H13 (located in China), H25, H26 and H27 (all located in Reunion) are individual and private, representing 0.54% of the total population for each.

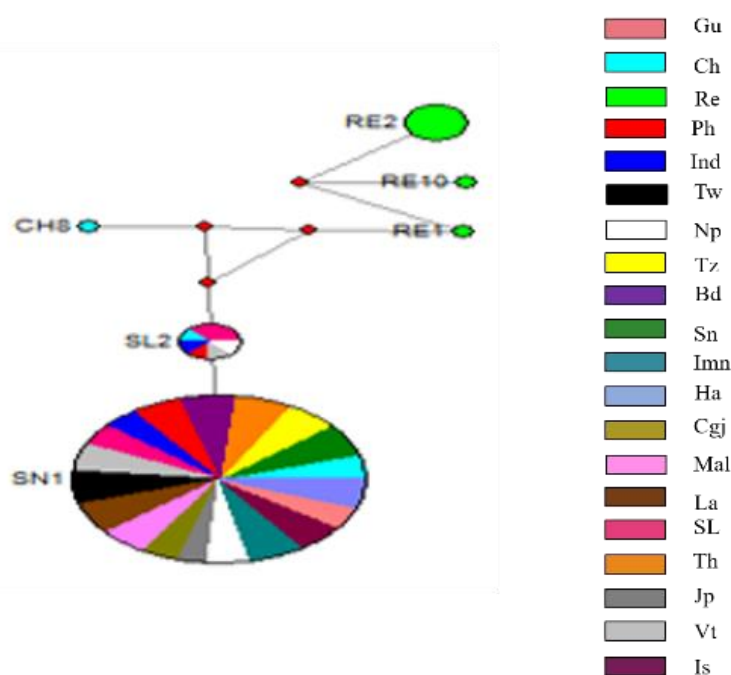


Figure 1. Haplotype network of *Zeugodacus cucurbitae*. The size of circles is proportional to the number of individuals in the haplotype or the number of haplotypes in the haplogroup.

3.2. Structuration and Genetic Distance

Analysis of genetic differentiation (F_{st}) (Table 4) shows

that only the F_{st} between the population of the meeting and all other populations are significant; between the population of Nepal and the populations of Senegal, Tanzania, Laos, and Hawaii. There is a very large difference between the Reunion

population and other populations. There is a slight difference between the populations of Nepal and those of Senegal, Tanzania, Laos, and Hawaii.

Table 4. *Fst values taken between populations.*

	Nepal	P-value	Reunion	P-value
Senegal	0.012	0.013	0.997	0.00
Tanzania	0.009	0.046	0.997	0.00
Thailand			0.997	0.00
Bangladesh			0.994	0.00
Sri Lanka			0.996	0.00
Indonesia			0.997	0.00
China			0.997	0.00
Vietnam			0.996	0.00
Philippines			0.997	0.00
Taiwan			0.994	0.00
Laos	0.09	0.039	0.996	0.00
Malaysia			0.996	0.00
Nepal			0.996	0.00

	Nepal	P-value	Reunion	P-value
Cambodia			0.996	0.00
Japan			0.996	0.00
Mariana Islands			0.997	0.00
Salomon Island			0.996	0.00
Guam			0.996	0.00
Hawaii	0,009	0,041	0.997	0.00

3.3. Genetic Distances

Considering two groups, with group 1 (G1) comprising all countries and group 2 (G2) composing all island sites.

The analysis of the genetic distance (Table 5) between the distance among populations within G1 is high and exceeds that observed within G2, which remains low. This suggest greater homogeneity among island populations and substantial differentiation among population in G1. Furthermore, the genetic distance between the two groups is higher than the within group distances. The COI gene displays marked genetic diversity between these groups.

Table 5. *Genetic distances between countries and islands.*

Distances		Intragroup distances	Intergroup distances	SD
Groups	G2	0.001		0.028
			0.213	0.017
	G1	0.338		0.000

If we consider the population of Reunion as a group, and all the other populations as another group, we obtain results in Table 6.

Table 6. *Genetic distances between 2 groups.*

Distances		Intragroup distances	Intergroup distances	SD
Groups	Reunion	0.003		0.001
			0.213	0.017
	Pays	0,001		0,000

If we consider as a group all the countries belonging to the same continent, we have five (5) groups which are: Africa, Asia, Oceania, Europe and the United States.

1. Intragroup distances

Genetic distances (Table 7) within Africa, Oceania, and US group are zero. The COI presents uniformity at the level of

these 3 groups. On the other hand, they are low between the populations of the Asian group and those of Europe.

Table 7. Intra-group distances between continents.

Groups	Distances	SD
Africa	0.000	0.000
Asia	0.001	0.000
Oceania	0.000	0.000
Europe	0.003	0.001

Groups	Distances	SD
US	0.000	0.000

2. Interpopulation distances

The results in Table 8 show high genetic distances between populations of the European group and those of other groups. This shows an isolation of the group from Europe compared to other groups. The genetic distance between the populations of the Africa group, the Oceania group and the United States group is zero. Similarly, the genetic distance between the populations of these 3 groups and those of the Asian group is very low.

Table 8. Inter-group genetic distances between continents.

Groups	Genetic distances	SD
Africa-Asia	0.001	0.000
Africa-Oceania	0.000	0.000
Asia-Oceania	0.001	0.000
Africa- United States	0.000	0.000
Asia- United States	0.001	0.000
Oceania- United States	0.000	0.000
Africa-Europe	0.954	0.080
Asia-Europe	0.955	0.080
Oceania-Europe	0.954	0.080
United States -Europe	0.954	0.080

3.4. Correlation Test Between Genetic Distance and Geographical Distance

The calculated p-value (0.012) is lower than the alpha significance level = 0.05 (Table 9), we must reject the null hypothesis H₀ and retain the alternative hypothesis H_a. There is indeed a correlation between geographical distance (Matrix A) and genetic differentiation (Matrix B). (Figures 2 & 3).

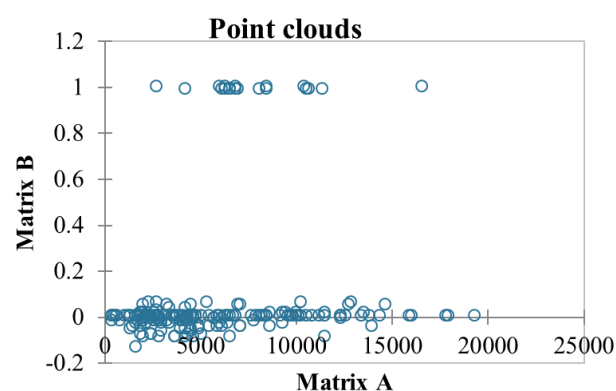


Figure 2. Correlation between geographical distances and F_{st} of *Z. cucurbitae*.

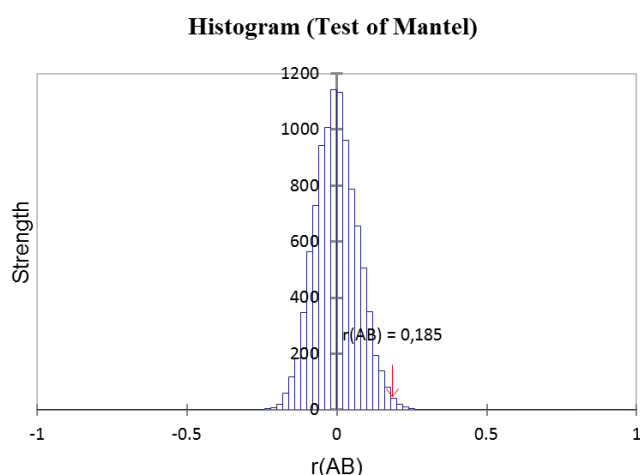


Figure 3. Mantel Pearson-type histogram of *Z. cucurbitae*.

Table 9. Test of Mantel results.

r (AB)	p-value	alpha
0.185	0.012	0.05

3.5. Structuration

The analysis of molecular variance by AMOVA of the 2 groups (Table 10) indicates that most of the molecular variance is due to the high average genetic differentiation between groups representing 99.81% of the total variation. The differentiation between individuals of the same population being very low is 0.19% of the total variation. However, the differentiation index (F_{st}) between groups is high and significant (0.99812).

Table 10. AMOVA test for 2 groups: Reunion and other populations.

Source of Variation	d.f	Sum of squares	Variance component	Variance percentage
Between groups	1	3107.501	164.39391Va	99.81
Between populations inside groups	18	5.006	-0.00345Vb	-0.00
Inside populations	162	50.103	0.30928Vc	0.19
Total	181	3162.610	164.69975	
Indices of fixation	F_{st} entre groupes = 0.99812 p value: 0.00000+-0.00000			

3.6. Demographic Evolution: D of Tajima and F_s of F_u

Results of Tables 11 and 12 indicate the values of Tajima's D and Fu's F_s significant within the populations. For all these populations, the values of D of Tajima and F_s of Fu are significantly negative, so we retain a demographic expansion of these regions.

Table 11. D Tajima of populations.

Populations	D of Tajima	P value
Bangladesh	-1.66706	0.03350
China	-1.72953	0.02990
Vietnam	-1.66706	0.02830
Nepal	-1.76663	0.01700
Reunion	-1.63600	0.04160

Table 12. F_s F_u of populations.

Populations	D of Tajima	P value
Bangladesh	2.84720	0.00160
Sri Lanka	-2.01642	0.04930
Vietnam	-1.34464	0.04530
Taiwan	-1.16394	0.03670
Nepal	-2.87184	0.00820

3.7. Phylogenetic Approach

The phylogenetic tree displays two clades (Figure 4). The first clade contains the haplotypes of Reunion. On the other hand, the second clade is composed of two subclades; one formed by haplotypes H8, H9, H10 from Sri Lanka, H13 from China, H16 from Vietnam and H18 from Nepal, and the larger one contains the remaining haplotypes. The Bayesian inference tree reflects the results of the haplotypes network.

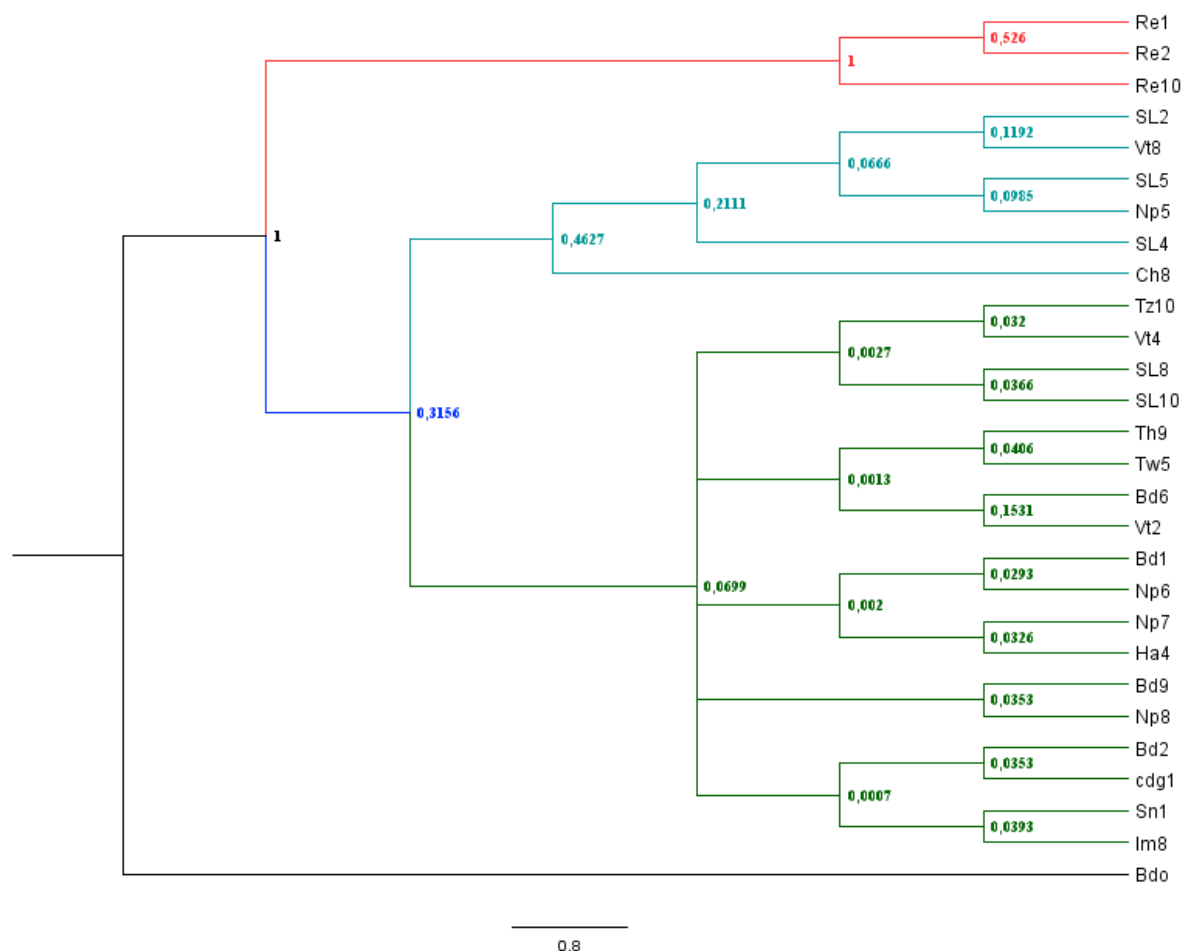


Figure 4. Phylogenetic tree of haplotypes by the Bayesian inference method.

4. Discussion

The objective of this study is to genetically characterize populations of *Zeugodacus cucurbitae*, watermelon pest insect, at the global level.

Our dataset contains 182 sequences. After sequence analysis, we found a high degree of polymorphism (52.70% on all sites, 96.15% of which are informative in parsimony and 3.84% are variable singletons). According to Gasparich *et al.* [31], the ancestor populations generally have high levels of genetic diversity unlike recent populations. Which suggests that our populations are close to the ancestor populations. The populations of Bangladesh, Sri Lanka, Vietnam, and Nepal all have high haplotypal (Hd) diversity and low nucleotide (Pi) diversity. This would indicate that these populations have a diversified genetic origin of ancestral populations which would have undergone a bottleneck followed by rapid population growth with an accumulation of mutations. This could be explained by the use of anthropogenic methods to control this pest. These results are somewhat in agreement with the work of Wu *et al.* [13] which suggests that western populations (Nepal, Bangladesh, Thailand,

Burma, and western China) exhibit greater diversity than eastern regions. Prabhakar *et al.* [12] state that the population expansion of this pest insect is an event related to hot post-Pleistocene climatic conditions with a small number of founder populations. On the other hand, all other populations have low nucleotide and haplotypic diversities; therefore these populations would have undergone a severe and prolonged demographic bottleneck which could be due to climate changes, interspecific competition because Denno *et al.* [32] concluded that interspecific competition significantly affected the distribution and abundance of phytophagous insects. This would lead to a genetic homogenization affirmed by the studies of Prabhakar *et al.* [12] with whom we have populations in common; they argue that the populations of *Zeugodacus cucurbitae* are homogeneous regardless of their geographical distribution. The set of populations presents strong haplotypic and nucleotide diversities; therefore an admixture, that is to say a mix of genes from different ancestral populations. This may result from migrations or gene flow between populations. The nucleotide changes made on all sequences allowed to detect 27 haplotypes with H1 being the dominant haplotype present in all populations except the Reunion population. The relationship between the mitochondrial haplo-

types was revealed by the network that shows 2 closely related haplogroups (SN1 and SL2), an individual haplotype at the level of Chinese propulsion and the three grouped haplotypes (H25, H26, H27) of the Reunion population distant from the latter. The unique H13 haplotype of the Chinese population would result from a recent mutation. This is not in agreement with the studies of Hu *et al.* [11] which estimate that the Chinese melon fly populations as a single and homogeneous phyletic unit. These three haplotypes within the Reunionese population reflect a greater intra-population diversity than between other countries. These results are supported by the intra-group distance within Reunion (0.003) which is greater than that between countries (0.001). Our results are in agreement with those of Jacquard *et al.* [33], who using ten microsatellites on two mitochondrial gene fragments found the existence of three genetically distinct groups of *Z. cucurbitae* at Reunion Island all clearly different from their African and Asian parents. The genetic differentiation index F_{st} between the Reunion population and other populations varies between 94 and 97%, which could be synonymous with a genetic isolation that could be due to geographical barriers, habitat differences that have limited gene flow. The strong genetic distance (0.213) between them testifies to this. The intra-population diversity within the meeting population could be a consequence of host preference due to specific intra competition. Our results are supported by the work of Virgilio *et al.* [8] who showed that the samples from the island of Reunion are genetically different from all other groups. The slight genetic differentiation between Nepal and the populations of Senegal, Tanzania, Laos and Hawaii would be justified by limited gene flow.

The Mantel Test reveals a correlation between geographical distance and F_{st} (bilateral p value < 0.05) (Figure 2). This finding is supported by the Mantel histogram (Figure 3) which presents a symmetric unimodal distribution. In other words, geographical distance would influence the genetic structuring of melon fly populations. This does not accord with the work of Wu *et al.* [13] in which there is no Reunion population and where geographical distances do not seem to influence the genetic structuring of the different melon fly populations. Molecular analysis (AMOVA) further confirms this genetic peculiarity of Reunion but also that there is no genetic differentiation between the other populations. This lack of genetic differentiation could be explained by the strong migratory capacity of *Z. cucurbitae*. The significant values of Tajima's D and Fu's F_s for the populations recorded in Tables 11 and 12 show a population growth that could be linked to climatic conditions. Globally at the level of each phylogenetic tree there is a clade that groups only individuals from the Reunion population. This would mean that this new variant of the Chinese population would come from the Reunionese population. This confirms the isolation by distance of the latter with the works of Virgilio *et al.* [8] which asserted a significant correlation between geographical distances and genetic distances. There is a clade that groups only the individuals from Vietnam and Bangladesh present in all the trees,

so the individuals of these populations would share mutations. The Bayesian inference tree presents more similarities with the haplotype network and reveals that melon fly populations can be subdivided into two main groups corresponding to populations coming from Reunion (1) and other continents (2). This is not in agreement with the work of Virgilio *et al.* [8] which subdivided these populations into five groups, including the African continent (1), Reunion (2), Central Asia (3), East Asia (4), and Hawaii (5).

5. Conclusion

This study, based on the genetic characterization of *Z. cucurbitae* populations using cytochrome oxidase I (COI), revealed the existence of isolation by distance and genetic structuring according to 2 groups: Reunion and other continents. An adaptation to the environment following a recent expansion of Reunion's population from an African or Asian melon fly population could be at the origin of this isolation. Furthermore, environmental differences have favoured genetic divergence.

Moreover, an intra-population structuring has been detected within the Reunionese population. This may be due to host preference in some populations given that the melon fly has a polyphagous diet.

Further studies would provide a better understanding of the structuring of *Z. cucurbitae* populations as distinct populations or subpopulations may have different responses to control methods such as pesticides or biological control agents.

Abbreviations

Re	Reunion
SL	Sri Lanka
Vt	Vietnam
Np	Nepal
Ch	China
Tz	Tanzania
Th	Thailand
Tw	Taiwan
Bd	Bangladesh
Ha	Hawaii
Im	Mariana Islands
Bdo	<i>Bactrocera dorsalis</i>

Author Contributions

Madeleine ivonne Mendy: Conceptualization, Writing - original draft, Investigation, Writing - review & editing, Methodology, Software

Toff ãe Diome: Resources, Writing - review & editing, Formal analysis

Mamecor Faye: Resources, Writing - review & editing, Visualization

Mback éSemb ène: Supervision, Validation

Conflicts of Interest

The authors declare no conflicts of interest.

References

- [1] Hdidier, C., Tlili, I., & Ilahy, R. (2020). Watermelon. In Nutritional Composition and Antioxidant Properties of Fruits and Vegetables. Elsevier. p. 515-531.
<https://doi.org/10.1016/B978-0-12-812780-3.00032-5>
- [2] Badji, H., & Coly, E. V. (2010). Gestion des Attaques de la Mouche des Fruits sur les Cultures de Cucurbitac ées au Moyen de Pesticides Naturels au S é n é gal. International Symposium on Urban and Peri-Urban Horticulture in the Century of Cities: Lessons, Challenges, Opportunités 1021, 421-426.
<https://doi.org/10.17660/ActaHortic.2014.1021.39>
- [3] Kunprom, C., & Pramual, P. (2017). Genetic structure and demographic history of the melon fly *Zeugodacus cucurbitae* (Coquillett) (Diptera: Tephritidae) in Thailand. Agr. For. Entomol, 20, 180-190. <https://doi.org/10.1111/afe.12242>
- [4] White, I. M., De Meyer, M. & Stonehouse, J. M. (2000) A review of native and introduced fruit flies (Diptera, Tephritidae) in the Indian Ocean islands of Mauritius, Réunion, and Seychelles. Proceedings, Indian Ocean Commission, Regional Fruit Fly Symposium (ed. by N. S. Price and S. I. Seewoosuruthun). Indian Ocean Commission/European Union, Flic en Flac, Mauritius. pp. 15-21.
- [5] Dhillon, M. K., Singh, R., Naresh, J. S., & Sharma, H. C. (2005). The melon fruit fly, *Bactrocera cucurbitae*: A review of its biology and management. Journal of Insect Science, 5(1), 40-56. <https://doi.org/10.1093/jis/5.1.40>
- [6] Bezzi, M. (1913). Indian Trypanoids [Trypetids](Fruit-Flies) in the Collection of the Indian Museum, Calcutta. Memoirs of The Indian Museum, 3, 53-168.
- [7] Nishida T, Bess HA (1957). Studies on the ecology and control of the melon fly *Dacus* (Strumeta) *cucurbitae* Coquillett (Diptera: Tephritidae). Hawaii Agricultural Experiment Station, University of Hawaii. Tech Bull 84: 12-29.
- [8] Virgilio, M., Delatte, H., Backeljau, T., & De Meyer, M. (2010). Macrogeographic population structuring in the cosmopolitan agricultural pest *Bactrocera cucurbitae* (Diptera: Tephritidae). Molecular Ecology, 19(13), 2713-2724.
<https://doi.org/10.1111/j.1365-294X.2010.04662.x>
- [9] Jacquard, C., Virgilio, M., David, P., Quilici, S., De Meyer, M., & Delatte, H. (2013). Population structure of the melon fly, *Bactrocera cucurbitae*, in Reunion Island. Biological Invasions, 15(4), 759-773. <https://doi.org/10.1007/s10530-012-032>
- [10] Delatte, H., De Meyer, M., & Virgilio, M. (2019). Genetic structure and range expansion of *Zeugodacus Cucurbitae* (Diptera: Tephritidae) in Africa. Bulletin of Entomological Research, 109(6), 713-722.
<https://doi.org/10.1017/S0007485319000026>
- [11] Hu Jian, H. J., Zhang, J. L., Nardi, F., & Zhang, R. J. (2008). Population genetic structure of the melon fly, *Bactrocera cucurbitae* (Diptera: Tephritidae), from China and Southeast Asia. 134, 319-324.
<https://doi.org/10.1007/s10709-007-9239-1>
- [12] Prabhakar, C. S., Mehta, P. K., Sood, P., Singh, S. K., Sharma, P., & Sharma, P. N. (2012). Population genetic structure of the melon fly, *Bactrocera cucurbitae* (Coquillett) (Diptera: Tephritidae) based on mitochondrial cytochrome oxidase (COI) gene sequences. Genetica, 140(1-3), 83-91.
<https://doi.org/10.1007/s10709-012-9660-y>
- [13] Wu, Y., McPherson, B. A., Wu, J., & Li, Z. (2012). Genetic relationship of the melon fly, *Bactrocera cucurbitae* (Diptera: Tephritidae) inferred from mitochondrial DNA. Insect Science, 19(2), 195-204.
<https://doi.org/10.1111/j.1744-7917.2011.01443.x>
- [14] Warot, S. F. (2018). Caract é risation mol é culaire et isolements reproducteurs chez des auxiliaires de lutte biologique. Mémoire. Université Paris sciences et lettres, Paris. 94p. HAL Id: hal-03236272 <https://ephe.hal.science/hal-03236272v1>
- [15] Avise, J. C., Arnold, J., Ball, R. M., Bermingham, E., Lamb, T., Neigel, J. E., Reeb, C. A., & Saunders, N. C. (1987). Intra-specific phylogeography: The Mitochondrial DNA Bridge Between Population Genetics and Systematics. Annual Review of Ecology and Systematics, 18(1), 489-522.
<https://www.jstor.org/stable/2097141>
- [16] Hebert, P. D. N., Cywinska, A., Ball, S. L., & deWaard, J. R. (2003). Biological identifications through DNA barcodes. Proceedings of the Royal Society of London. Series B: Biological Sciences, 270(1512), 313-321.
<https://doi.org/10.1098/rspb.2002.2218>
- [17] Ndiaye, M. R. (2018). Evolution phylog éographique et structure g énéétique des populations de *Sitophilus zeamais* (Coleoptera, Curculionidae), ravageur du ma ï stocké en Afrique de l'Ouest [Phdthesis, Université Cheikh Anta DIOP de Dakar, S é n é gal ; Faculté des Sciences et Techniques; Département de Biologie Animale]. 178p.
<https://hal.science/tel-02116356>
- [18] Sabir, J. S. M., Rabah, S., Yacoub, H., Hajrah, N. H., Atef, A., Al-Matary, M., Edris, S., Alharbi, M. G., Ganash, M., Mahyoub, J., Al-Hindi, R. R., Al-Ghamdi, K. M., Hall, N., Bahieldin, A., Kamli, M. R., & Rather, I. A. (2019). Molecular evolution of cytochrome C oxidase-I protein of insects living in Saudi Arabia. PLoS ONE, 14(11), 1-18.
<https://doi.org/10.1371/journal.pone.0224336>
- [19] NDONG, A., THIAW, C., Diallo, B., SARR, M., DIOME, T., KANE, M., & SEMBENE, M. (2015). Barcoding: Comparison of Variation Degree of COI and Cytochrome b Mitochondrial Markers in Two Species Primary Maize Pests (*Sitophilus zeamais* and *Sitophilus oryzae*). International Journal of Sciences: Basic and Applied Research, 20(1), 373-393.
<https://www.gssrr.org/index.php/JournalOfBasicAndApplied/article/view/3420>

- [20] Avise, J. C. (2000). *Phylogeography: The history and formation of species*. Harvard university press. Cambridge, MA. 441p.
- [21] Saccone, C., De Giorgi, C., Gissi, C., Pesole, G., & Reyes, A. (1999). Evolutionary genomics in Metazoa: The mitochondrial DNA as a model system. *Gene*, 238(1), 195-209. [https://doi.org/10.1016/S0378-1119\(99\)00270-X](https://doi.org/10.1016/S0378-1119(99)00270-X)
- [22] Hall, T. A. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic acids symposium series*, 41(41), p 95-98.
- [23] Librado, P., & Rozas, J. (2009). DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. *Bioinformatics*, 25(11), 1451-1452. <https://doi.org/10.1093/bioinformatics/btp187>
- [24] Tamura, K., Stecher, G., Peterson, D., Filipinski, A., & Kumar, S. (2013). MEGA6: Molecular evolutionary genetics analysis version 6.0. *Molecular biology and evolution*, 30(12), 2725-2729. <https://doi.org/10.1093/molbev/mst197>
- [25] Nei, M. (1973). Analysis of Gene Diversity in Subdivided Populations. *Proceedings of the National Academy of Sciences*, 70(12), 3321-3323. <https://doi.org/10.1073/pnas.70.12.3321>
- [26] Nei, M. (1977). F - statistics and analysis of gene diversity in subdivided populations. *Annals of Human Genetics*, 41(2), 225-233. <https://doi.org/10.1111/j.1469-1809.1977.tb01918.x>
- [27] Bandelt, H. J., Forster, P., Röhl, A., (1999). Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution*, 16(1), 37-48, <https://doi.org/10.1093/oxfordjournals.molbev.a026036>
- [28] Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, 10(3), 564-567. <https://doi.org/10.1111/j.1755-0998.2010.02847.x>
- [29] Huelsenbeck, J. P., & Ronquist, F. (2001). MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics*, 17(8), 754-755.
- [30] Rambaut, A., & Drummond, A. J. (2012). Figtree, a graphical viewer of phylogenetic trees. V. 1.4. 0. Available from <http://tree.bio.ed.ac.uk/software/figtree/>
- [31] Gasparich, G. E., Silva, J. G., Han, H.-Y., Mcpherson, B. A., Steck, G. J., & Sheppard, W. S. (1997). Population genetic structure of Mediterranean fruit fly (Diptera: Tephritidae) and implications for worldwide colonization patterns. *Annals of the Entomological Society of America*, 90(6), 790-797. <https://doi.org/10.1093/aesa/90.6.790>
- [32] Denno, R. F., McClure, M. S., & Ott, J. R. (1995). Interspecific interactions in phytophagous insects: Competition reexamined and resurrected. *Annual review of entomology*, 40(1), 297-331.
- [33] Jacquard, C., Virgilio, M., Quilici, S., De Meyer, M., & Delatte, H. (2012). Spatial scales of genetic structuring in *Bactrocera cucurbitae* (Diptera, Tephritidae): Population structure on la Réunion. <https://agritrop.cirad.fr/563683>